

MAGNESIUM BINDING TO CALCIUM-BINDING PROTEINS:  
A REGULATORY FUNCTION?

Torbjörn Drakenberg, Sture Forsén, Eva Thulin  
and Ming-Daw Tsai<sup>1</sup>

Division of Physical Chemistry 2  
University of Lund  
Lund, Sweden

I. INTRODUCTION

It is generally accepted that  $Mg^{2+}$ -ions serve a structural role in binding to the "Ca-Mg" sites of troponin C [1]. It is thus believed that  $Mg^{2+}$  binding to TnC ensures that the troponin complex stays intact also in the resting, low  $Ca^{2+}$ , state of the muscle. The two weaker  $Ca^{2+}$  sites in TnC, the regulatory sites, are normally assumed not to bind  $Mg^{2+}$ -ions under physiological conditions. For calmodulin there is no clear agreement regarding the  $Mg^{2+}$  binding, reported values ranging from 0 to  $10^4 M^{-1}$  [2].

We have previously used  $^{43}Ca$  NMR to study mainly the kinetics in the calcium binding to TnC and CaM [3]. These studies show that for both proteins there are two classes of sites with different exchange dynamics for the  $Ca^{2+}$ -ions. Studies of the tryptic fragment show that the C-terminal half contains the two stronger sites with slower  $Ca^{2+}$ -ion exchange and the N-terminal half contains the two weaker sites with faster exchange rates [4]. We have now used the same approach to study  $Mg^{2+}$ -ion binding to TnC and CaM by means of  $^{25}Mg$  NMR. These studies show undoubtedly that there are two  $Mg^{2+}$  binding sites in each half of the two proteins. Furthermore, we have used a Phenyl-Sepharose column to investigate if CaM,

<sup>1</sup>Permanent address: Department of Chemistry, The Ohio State University, Columbus, Ohio.

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in which  
 $Mg^{2+}$ -ions  
(Ca)<sub>2</sub>(Mg)  
(Ca)<sub>4</sub>CaM

II. EXPERIMENTAL

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III. RESULTS

A. Troponin C

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B. Calmodulin

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in which some of the  $\text{Ca}^{2+}$ -ions have been replaced by  $\text{Mg}^{2+}$ -ions, will still bind. These studies show that  $(\text{Ca})_2(\text{Mg})_2\text{CaM}$  binds to the column in the same way as  $(\text{Ca})_4\text{CaM}$ .

## II. EXPERIMENTAL

The elution experiments with Phenyl-Sepharose columns were made using a reversed salt gradient.

All NMR experiments were performed on a Nicolet 360 WB spectrometer at 22.15 MHz for  $^{25}\text{Mg}$  and 24.34 MHz for  $^{43}\text{Ca}$  using a homemade probe.

## III. RESULTS AND DISCUSSION

### A. Troponin C

The temperature dependence in the  $^{43}\text{Ca}$  NMR line width for a sample containing 1 mM  $\text{TR}_1\text{C}$  from TnC and an excess of  $\text{Ca}^{2+}$  is that typical for intermediate exchange and similar to TnC itself. Adding  $\text{Mg}^{2+}$ -ions results in a decrease in the  $^{43}\text{Ca}$  NMR line width. Considering reported binding constants for  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$ -ions to TnC [1] and its fragments our results are only in agreement with a model where  $\text{Mg}^{2+}$ -ions can bind to site(s) different from the  $\text{Ca}^{2+}$  binding sites and that this ion binding will modulate the dynamics in the binding of the  $\text{Ca}^{2+}$ -ions.

$^{25}\text{Mg}$  NMR studies of  $\text{Mg}^{2+}$  binding to TnC and its tryptic fragments show that there are two strong  $\text{Mg}^{2+}$ -sites in the C-terminal half of the protein ( $\text{TR}_2\text{C}$ ) with slow exchange dynamics for the  $\text{Mg}^{2+}$ -ions. In the N-terminal half of the protein ( $\text{TR}_1\text{C}$ ) there are  $\text{Mg}^{2+}$  binding sites that are significantly weaker than those in  $\text{TR}_2\text{C}$ . The off-rate of the  $\text{Mg}^{2+}$ -ions from these sites are much faster ( $\sim 10^3 \text{ s}^{-1}$ ) than for  $\text{TR}_2\text{C}$ . It is, however, at present not clear how many if any of these sites are the same as the  $\text{Ca}^{2+}$ -sites.

### B. Calmodulin

The two stronger (slowly exchanging)  $\text{Ca}^{2+}$  sites are located in the C-terminal half of the protein whereas the two weaker (faster exchanging) sites are in the N-terminal half [3].  $^{25}\text{Mg}$  NMR data show that  $\text{Mg}^{2+}$ -ions bind to both halves of CaM. It is also evident from the concentration dependence of the line width that  $\text{TR}_1\text{C}$  binds  $\text{Mg}^{2+}$ -ions stronger by a factor

of 10 than TR<sub>2</sub>C. We have thus found that Mg<sup>2+</sup>-ions have a reversed site preference as compared to Ca<sup>2+</sup>-ions, however, the same as the lanthanides. The temperature dependence in the width of the <sup>25</sup>Mg line has been used to calculate the exchange rate of the Mg<sup>2+</sup>-ions resulting in an off-rate of Mg<sup>2+</sup>-ions from TR<sub>1</sub>C of 2,000 - 3,000 s<sup>-1</sup>, whereas the one from TR<sub>2</sub>C is not well defined from our experiments, however, faster than for TR<sub>1</sub>C.

The reasonably strong binding of Mg<sup>2+</sup> to TR<sub>1</sub>C and to the N-terminal half in intact CaM show that Mg<sup>2+</sup> will compete favourably with Ca<sup>2+</sup> for these sites, and in the non-activated cell (low calcium) these sites will be saturated by magnesium to well over 50 %, maybe 90 %. This binding can have several implications for the mechanism of CaM modulation. To test if partially Mg<sup>2+</sup> loaded CaM may bind to hydrophobic sites we have studied the binding of CaM to a Phenyl-Sepharose column at different Mg<sup>2+</sup> and Ca<sup>2+</sup> concentrations. In the absence of Mg<sup>2+</sup> a [Ca<sup>2+</sup>]/[CaM] ratio of 4 is needed for binding to the column. When 10 mM Mg<sup>2+</sup> was present a [Ca<sup>2+</sup>]/[CaM] ratio of 2 was however sufficient to bind all CaM to the column. This shows that two of the four Ca<sup>2+</sup>-ions in CaM can be replaced by Mg<sup>2+</sup>-ions and the conformation of CaM still is the right one for binding to Phenyl-Sepharose and presumably to hydrophobic drugs and target enzymes. Whether or not (Ca)<sub>2</sub>(Mg)<sub>2</sub>CaM will also modulate the target enzyme is another question not addressed in this work.

#### REFERENCES

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